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EFFECT OF NAPHTHALENE ACETIC ACID (NAA) ON IN VITRO ROOTING OF REGENERATED MICROSHOOTS OF GROUNDNUT (*ARACHIS HYPOGAEA* L.)

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ABSTRACT

Successful utilization of tissue culture for the improvement and conservation of groundnut will be made possible only with an efficient shoot proliferation, formation of well-developed root system in microshoots, successful acclimatization and final establishment in field. A regeneration protocol ensuring a high frequency rooting of microshoots derived from embryonic axes of four groundnut genotypes was achieved using naphthalene acetic acid (NAA). Microshoots of 3cm length were subcultured on Murashige and Skoog (MS) medium plus 30% sucrose, supplement with 1.0 or 1.5mg/L NAA and compared with a hormone free medium (control). Rooting was significantly influenced by the presence and concentration of NAA. Microshoots subcultured in medium supplemented with 1mg/L NAA rooted within 2weeks of subculture, producing the highest number of roots/plantlet and root-induction frequency. However, increasing the concentration of the auxin suppressed root length with the hormone free medium having the longest roots. Significant genotypic effect was also observed, with Samnut 22 having more number of roots/plantlet and root-induction frequency.

Keywords: groundnut. microshoots. in vitro rnttina. NAA

INTRODUCTION

The potential of tissue culture for conservation and genetic improvement of groundnut is well recognized (Gagliardi *et al.* 2002, 2003; Mckentiy *et al.* 1995; Swathi *et al.* 2006; Venkatachalam *et al.* 2000). However, successful utilization of tissue culture will be made possible only with an efficient shoot proliferation, formation of well developed root system in microshoots, successful acclimatization and final establishment in field. This under scores the need to established an efficient root induction protocol for successful field establishment. Mostly, *in vitro* rooting of microshoots has been achieved using media fortified with auxins either singly or in combination with cytokinins. Several rooting media have been employed to induce rooting in groundnut. Rooting was induced in groundnut on medium fortified with 2mg/L NAA (Nazir *et al.* 2011) and a combination of indole butyric acid (IBA) and kinetin (KIN) (Venkatachalam and Jayabalan, 1997). While Gagliardi *et al.* (2000) were able to obtain rooting in groundnut on hormone free medium. Generally, groundnut response to tissue culture is strongly influenced by the plant genotype (Mroginsky *et al.* 1981; Banerjee *et al.* 2007). Verma *et al.* (2009) were able to induce rooting in four genotypes of groundnut on MS medium fortified with 1mg/L NAA, while a combination of 1mg/L NAA and 0.5mg/L IBA was required for rooting the fifth genotype.

In view of the genotypic influence in groundnut response to tissue culture, this study was carried with the objective of establishing an efficient root induction protocol applicable to a wide range of groundnut genotypes.

MATERIALS AND METHODS

Microshoots of 3cm length (Figure 1) were subcultured on MS medium (Murashige and Skoog, 1962) plus 30% sucrose and supplement with 1.0 or 1.5mg/L NAA and compared with a hormone free medium (control). The pH of the media was adjusted to 5.8 before autoclaving for 15min at 121°C. Ten test tubes were used per treatment and these were laid out in a completely randomized design replicated three times. Cultures were maintained in growth chamber at 26±2°C under 16hr light/8 hr dark photoperiod for five weeks. Well rooted plantlets were washed under running tap water to remove the medium from the roots. Root induction frequency, number of roots/plantlet and root length were taken for each treatment. Thereafter the plantlets were transferred to sterilized riverside sand and covered with transparent plastic bags to retain humidity and maintained at room temperature for 7 days before transferred to the screen house. Data collected were subjected to analysis of variance and means separated using DMRT.



Figure 1: Regenerated microshoot of groundnut on MS medium+15mg/L BAP

RESULTS AND DISCUSSION

The presence and concentration of NAA significantly influenced rooting in groundnut. Rooting was observed within two weeks in responsive microshoots. Medium supplemented with 1mg/L NAA recorded the highest root induction frequency (82.42%) compared to 1.5mg/L NAA treatment (63.55%). Similarly, 1.5mg/L NAA treatment produced higher root induction frequency compared to the hormone free medium (10.42%) which recorded the least (Table 1). This result suggests that NAA is essential for high rooting frequency in groundnut microshoots as minimal rooting was observed in its absences. It has been established that auxin stimulates lateral root initiation by activating quiescent pericycle cells to initiate division and then expansion which facilitate lateral root emergence (Fukaki and Tasaka, 2009). Therefore appropriate synthesis, signaling and transport of auxin are all required for root formation (Peret *et al.*, 2009). Root induction frequency obtained at 1mg/L was higher than the frequency reported by McKently *et al.* (1990) at the same concentration. On the contrary, Gagliardi *et al.* (2000) reported high development of roots on MS hormone free medium. The effect of NAA on number of roots/plantlet was significant. Medium supplemented with 1mg/L NAA and 1.5mg/L NAA produced comparable higher number of roots/plantlets (9.25 & 9.42) compared to hormone free medium (Table 1; Figure 2). Auxin controls cell division, cell elongation and certain stages of differentiation (Davies, 2004) which could have increased the number of roots/plantlet. Lewis *et al.* (2011), reported that auxin increases the number of

lateral roots. The value obtained in our study is higher than that reported by McKently *et al.* (1990) on basal medium supplemented with 1mg/L NAA.

However, the presence and concentration of NAA in the medium decreased the root length. Hormone free medium produced the longest root length (7.35cm) among the treatments (Table 1). Taiz and Ziegler (2003) reported that although plants may require auxin for root induction but root growth is inhibited at higher concentration. Kollmeier *et al.* (2000) reported that root elongation phase is very sensitive to auxin concentration and it is inhibited by high concentration of auxin in the rooting medium. Auxin can increase the rate of ethylene biosynthesis (Riov and Yang, 1989). Therefore it could be possible that high concentration of NAA inhibited root elongation through the inducement of ethylene biosynthesis, which is inhibitory to root growth.

Significant difference ($P < 0.05$) was observed among the groundnut genotypes in response to the NAA treatment. Samnut 22 and Samnut 21 recorded the highest root induction frequency of 60.00 and 55.19% respectively, and were significantly higher than Samnut 23 and Samnut 10 (Fig 3). Similarly, effect of NAA on number of roots per plantlets was significant, with Samnut 22, Samnut 23 and Samnut 21 producing more number of roots/plantlets than Samnut 10 (Fig 4). However, Samnut10 and Samnut21 produced the longest root lengths compare to the other genotypes (Fig 4). This result confirms earlier reports that rooting in groundnut is strongly influenced by the plant genotype (Venkatachalam and Jayabalan, 1997; Verma *et al.*, 2009).

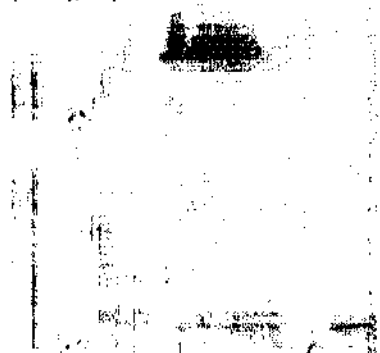


Fig 2: Plantlets after 4weeks on rooting media (MS supplemented with 1.5, 1mg/L NAA and control)

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